

Tetrahedron Letters 43 (2002) 379-382

TETRAHEDRON LETTERS

Syntheses of (-)-(7S)- and (+)-(7R)-K252a dimension

Kazuhiko Tamaki, Elliott W. D. Huntsman, Dejah T. Petsch and John L. Wood*

Sterling Chemistry Laboratories, Department of Chemistry, Yale University, New Haven, CT 06520-8107, USA Received 7 August 2001; revised 8 November 2001; accepted 13 November 2001

Abstract—This letter describes syntheses of (-)-(7S)-and (+)-(7R)-K252a dimers wherein a convergent bis-indole-*N*-glycosidic coupling step was used for the resolution of the C(7) substituted (\pm) -aglycon. Dimerization of the derived monomers was achieved via olefin methathesis. © 2002 Elsevier Science Ltd. All rights reserved.

K252a (1) (Fig. 1),¹ isolated originally from the culture broth of Actinomadura by Sezaki and then reisolated from Nocardiopsis by Kase in a screen for antagonists of Ca²⁺ mediated signaling, has been implicated in the inhibition of several tyrosine kinases (Trks) and has been shown to be a potent PKC inhibitor.² Trk mediated PDGF signal transduction is potentially a novel therapeutic inroad for the treatment of human gliomas and follows the general motif of receptor tyrosine kinases (i.e. dimerization and autophosphorylation).³ Based on recent work demonstrating that dimeric natural product probes function particularly efficiently as tools for chemical biology,⁴⁻⁶ we became curious as to whether multivalent analogs of K252a would have a selective influence on receptor-type kinases. Thus, we initiated an effort to prepare K252a analogs poised for subsequent conversion to dimeric or higher valent species. Recently, efforts to access novel K252a analogs have resulted in a convergent route to C7 and C2' alkylated K252a analogs.^{7a-d} Herein, we describe a synthesis of (-)-(7S)-and (+)-(7R)-K252a dimers (2 and 3) utilizing olefin methathesis. The requisite monomers were prepared using an expansion of our strategy for the construction of C7 alkylated K252a analogs.^{7b,c}

The synthesis of (\pm) -aglycon 11 is outlined in Scheme 1. Thus, alkylation of known imine 4 with 8-bromo-1octene according to O'Donnel's procedure followed by hydrolysis of the imine afforded ethyl 2-amino-9decenoate hydrochloride (5) in 72% overall yield.8 A two-step reductive amination utilizing 3,4-dimethoxybenzaldehyde and NaBH₄ gave DMB (3,4-dimethoxybenzyl) protected amine 6 in 81% yield. Acylation with ethyl hydrogen malonate under standard coupling conditions (DCC/DMAP) afforded 7 in 92% yield. Dieckmann cyclization (NaOEt/EtOH) followed bv decarboalkoxylation in wet CH₃CN gave N-DMB-protected tetramic acid 8 (84%, two steps). Subsequent diazotransfer with 4-nitrobenzenesulfonyl azide in the presence of Et₃N smoothly gave diazolactam 9 in 93% vield.⁹ According to previous protocols, 9 and biindole 10 were treated with $Rh_2(OAc)_4$ in degassed pinacolone under reflux for 8 h. Successive C-H insertion, electrocyclization and aromatization gave the desired N-DMB



Figure 1.

^{*} Corresponding author. Fax: 203-432-6104; e-mail: john.wood@yale.edu

^{0040-4039/02/\$ -} see front matter @ 2002 Elsevier Science Ltd. All rights reserved. PII: S0040-4039(01)02166-9



Scheme 1.

protected aglycon (\pm)-11 in 33% yield (57% based on recovered 10).^{7a} With 11 in hand we explored the one-step acid catalyzed bis-indole-*N*-glycosidic coupling with enantioenriched 12a-12d.^{7a,10} Slow addition (24 h) of 12a-12d (a solution in 1,2-dichloroethane) to 11 and CSA (0.1 equiv.) in 1,2-dichloroethane followed by an additional 48 h at reflux gave a 2:2:1:1 diastereomeric mixture of methathesis precursors (13-16). Following isolation of the individual diastereomers by careful preparative TLC,¹¹⁻¹⁴ the initial regiochemical assignment for 13-16 was made based on ¹H NMR analysis. Subsequent assignment of the relative stereochemistry to the major regioisomers (-)-13 and (+)-14 was made by comparison to previously prepared C7methyl-6-*N*-DMB-K252a^{7b,c} and C7-benzyl-6-*N*-DMB-K252a^{7c} analogs. The relative stereochemistry of (+)-15 and (-)-16 remains ambiguous due to a lack of comparison data.

Treatment of (-)-13 with Grubbs' catalyst (Scheme 2)¹⁵ (8 mol%) at room temperature in CH_2Cl_2 for 43 h gave the desired dimer as a mixture of *E* and *Z* isomers.¹⁶ Subsequent hydrogenation with 5% Rh/Al₂O₃ afforded (7*S*)-6-*N*-DMB-dimer 17 in 87% overall yield. Final



DMB deprotection was carried out with TFA in the presence of anisole to provide the (-)-(7S)-K252a dimer (2) in 50% yield.¹⁷ An identical reaction sequence utilizing (+)-**14** furnished the corresponding (+)-(7R)-K252a dimer (3) in 49% overall yield.¹⁸

In conclusion, the first total synthesis of a C7 linked dimer of K252a has been completed. The synthesis expands the scope of our aglycon synthesis and establishes the feasibility of using cyclofuranosylation with enentioenriched carbohydrate precursors as a means to resolve racemic aglycons. Evaluation of the biological activity of these dimers is underway and has already established them to be potent kinase inhibitors.

Acknowledgements

We acknowledge the support of this work by Bristol– Myers Squibb, Eli Lilly, Glaxo–Wellcome, Merck, Pfizer, Yamanouchi, and Zeneca through their Faculty Awards Programs and the Camille and Henry Dreyfus Foundation for a Teacher–Scholar Award. J.L.W. is a fellow of the Alfred P. Sloan Foundation. K.T. thanks Sankyo Co., Ltd. for research support and J. Brad Shotwell for helpful comments during the preparation of the manuscript.

References

- Sezaki, M.; Sasaki, T.; Nakazawa, T.; Takeda, U.; Iwata, M.; Watanabe, T.; Koyama, M.; Kai, F.; Shomura, T.; Kojima, M. *J. Antibiot.* **1985**, *38*, 1437–1439.
- (a) Kase, H.; Iwahashi, K.; Matsuda, Y. J. Antibiot. 1986, 39, 1059–1065; (b) Nakanishi, S.; Matsuda, Y.; Iwahashi, K.; Kase, H. J. Antibiot. 1986, 39, 1066–1071; (c) Yasuzawa, T.; Iida, T.; Yoshida, M.; Hirayama, N.; Takahashi, M.; Shirahata, K.; Sano, H. J. Antibiot. 1986, 39, 1072–1078.
- Chin, L. S.; Murray, S. F.; Zitnay, K. M.; Rami, B. Clin. Cancer. Res 1997, 3, 771–776.
- Indeed, several dimeric natural product-like inhibitors have been designed and synthesized, including FK506⁴, cyclosporin A⁵, and 1α,25-dihydroxyvitamin D₃;⁶ (a) Spencer, D. M.; Wandless, T. J.; Schreiber, S. L.; Crabtree, G. R. *Science* 1993, 262, 1019–1024; (b) Pruschy, M. N.; Spencer, D. M.; Kapoor, T. M.; Miyake, H.; Crabtree, G. R.; Schreiber, S. L. *Chem. Biol.* 1994, *1*, 163–172; (c) Diver, S. T.; Schreiber, S. L. *J. Am. Chem. Soc.* 1997, 119, 5106–5109.
- Belshaw, P. J.; Spencer, D. M.; Crabtree, G. R.; Schreiber, S. L. Chem. Biol. 1996, 3, 731–738.
- Sestelo, J. P.; Mouriño, A.; Sarandeses, L. A. J. Org. Chem. 2000, 65, 8290–8296.
- In this study, the choices of linker length and position of attachment (i.e. C-7) were based soley on synthetic convenience, see: (a) Wood, J. L.; Stoltz, B. M.; Dietrich, H.-J.; Pflum, D. A.; Petsch, D. T. J. Am. Chem. Soc. 1997, 119, 9641–9651; (b) Wood, J. L.; Petsch, D. T.; Stoltz, B. M.; Hawkins, E. M.; Elbaum, D.; Stover, D. R. Synthesis 1999, 1529–1533; (c) Petsch. D. T. Ph.D. Thesis, Yale

University, 1999; (d) Tamaki, K.; Shotwell, J. B.; White, R. D.; Drutu, I.; Petsch, D. T.; Nheu, T. V.; He, H.; Hirokawa, Y.; Maruta, H.; Wood, J. L. *Org. Lett.* **2001**, 3, 1689–1692.

- O'Donnell, M. J.; Wojciechowski, K.; Ghosez, L.; Navarro, M.; Sainte, F.; Antoine, J.-P. Synthesis 1984, 313–315.
- Rector, D. L.; Harmon, R. E. J. Org. Chem. 1966, 31, 2837–2841.
- The four diasteromers (12a-12d) were all derived from alcohol 18 (92% ee) and lead to identical cyclofulanosylation products. See Ref. 7a.



- 11. (-)-(7S)-(1-Octenyl)-N-DMB-K252a (13): a pale yellow powder; $[\alpha]_D^{20}$ –36.0 (c 0.30, CHCl₃); IR (thin film/NaCl) 3298 (br w), 2930 (m), 2854 (w), 1733 (m), 1673 (s), 1646 (s), 1585 (m), 1515 (m), 1459 (s), 1392 (m) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.51 (d, J = 7.9 Hz, 1H), 8.09 (d, J=7.3 Hz, 1H), 8.00 (d, J=8.2 Hz, 1H), 7.81 (d, J=8.1 Hz, 1H), 7.51 (app t, J=7.8 Hz, 1H), 7.43 (app t, J = 7.8 Hz, 1H), 7.36–7.26 (m, 2H), 7.17 (dd, J = 5.0, 7.6 Hz, 1H), 7.11 (d, J=1.8 Hz, 1H), 7.01 (dd, J=1.8, 8.3 Hz, 1H), 6.92 (d, J=8.3 Hz, 1H), 5.60 (ddt, J=10.2, 17.0, 6.8 Hz, 1H), 5.34 (app t, J=3.2 Hz, 1H), 5.34 (d, J = 15.1 Hz, 1H), 5.27 (s, 1H), 4.80 (m, 1H), 4.76 (m, 1H), 4.42 (d, J=15.1 Hz, 1H), 4.01 (s, 3H), 3.77 (s, 3H), 3.76 (s, 3H), 3.49 (dd, J=7.6, 14.2 Hz, 1H), 2.52 (m, 1H), 2.41 (m, 1H), 2.23 (s, 3H), 2.20 (m, 1H), 1.82–1.73 (m, 2H), 1.12-0.92 (m, 7H), 0.54 (m, 1H); ¹³C NMR (100 MHz, $CDCl_3$): δ 173.8, 169.8, 149.0, 148.2, 140.3, 138.9, 137.0, 134.2, 130.4, 128.9, 126.4, 125.6, 125.0, 124.5, 124.0, 122.6, 121.9, 120.6, 120.1, 119.6, 119.3, 116.7, 114.8, 114.2, 113.9, 111.0, 110.7, 107.3, 98.9, 85.3, 84.7, 60.0, 55.8, 55.8, 53.7, 43.6, 42.2, 33.5, 29.2, 29.0, 28.6, 28.5, 22.8, 21.6; HRMS (FAB) m/z 728.3339 [calcd for C₄₄H₄₆N₃O₇ (M+H) 728.3336].
- 12. (+)-(7R)-(1-Octenyl)-N-DMB-K252a (14): a pale yellow powder; $[\alpha]_{D}^{20}$ +74.8 (c 0.21, CHCl₃); IR (thin film/NaCl) 3485 (br w), 3304 (br w), 3069 (w), 2998 (m), 2928 (s), 2854 (m), 1732 (s), 1673 (s), 1585 (m), 1515 (s), 1452 (s), 1392 (s) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.51 (d, J=7.8 Hz, 1H), 8.12 (d, J=7.6 Hz, 1H), 8.01 (d, J=8.5 Hz, 1H), 7.80 (d, J=8.3 Hz, 1H), 7.51 (app t, J = 7.4 Hz, 1H), 7.43 (app t, J = 7.5 Hz, 1H), 7.36–7.27 (m, 2H), 7.15 (dd, J = 5.1, 7.3 Hz, 1H), 7.12 (d, J = 1.8Hz, 1H), 7.02 (dd, J=1.8, 8.2 Hz, 1H), 6.93 (d, J=8.2Hz, 1H), 5.61 (ddt, J=10.3, 17.1, 6.8 Hz, 1H), 5.39 (s, 1H), 5.36 (m, 1H), 5.34 (d, J=15.2 Hz, 1H), 4.81 (m, 1H), 4.76 (m, 1H), 4.42 (d, J = 15.2 Hz, 1H), 4.02 (s, 3H), 3.78 (s, 3H), 3.77 (s, 3H), 3.51 (dd, J=7.3, 14.2 Hz, 1H), 2.54-2.35 (m, 2H), 2.36 (dd, J=5.1, 14.2 Hz, 1H), 2.18 (s, 3H), 1.82–1.75 (m, 2H), 1.12–0.83 (m, 7H), 0.50 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 174.2, 169.8, 149.2, 148.3, 140.1, 139.0, 137.2, 134.7, 130.3, 128.8, 126.9, 125.8, 125.0, 124.4, 124.3, 123.3, 122.4, 120.6, 120.3, 120.2, 117.1, 114.9, 114.0, 113.9, 111.0, 111.0, 107.3, 98.5, 85.0, 84.3, 58.7, 55.9, 55.8, 54.2, 43.6, 41.5, 33.6, 29.2, 29.1, 28.8, 28.6, 22.8, 21.6; HRMS (FAB) m/z 728.3339 [calcd for C₄₄H₄₆N₃O₇ (M+H) 728.3336].

- 13. (+)-Indolocarbazole 15 (isomer 1): a pale yellow powder; $[\alpha]_{D}^{20}$ +68.7 (c 0.31, CHCl₃); IR (thin film/NaCl) 3485 (br m), 3333 (br m), 3056 (m), 2999 (m), 2928 (s), 2854 (m), 1732 (s), 1671 (s), 1585 (m), 1514 (s), 1451 (s), 1400 (s) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.76 (d, J = 8.0Hz, 1H), 8.14 (d, J=7.8 Hz, 1H), 7.94 (d, J=8.6 Hz, 1H), 7.88 (d, J = 8.5 Hz, 1H), 7.51 (app t, J = 7.7 Hz, 1H), 7.44 (app t, J=7.8 Hz, 1H), 7.35–7.27 (m, 2H), 7.16 (dd, J=5.0, 7.3 Hz, 1H), 7.10 (d, J=1.9 Hz, 1H), 7.01 (dd, J=1.9, 8.2 Hz, 1H), 6.91 (d, J=8.1 Hz, 1H), 5.60 (ddt, J = 10.2, 17.0, 6.8 Hz, 1H), 5.34 (d, J = 14.9 Hz, 1H), 5.30 (app t, J=3.7 Hz, 1H), 5.26 (s, 1H), 4.81 (m, 1H), 4.76 (m, 1H), 4.42 (d, J=14.9 Hz, 1H), 4.00 (s, 3H), 3.77 (s, 3H), 3.75 (s, 3H), 3.49 (dd, J = 7.3, 13.8 Hz, 1H), 2.53 (m, 1H), 2.42 (m, 1H), 2.26 (m, 1H), 2.23 (s, 3H), 1.82–1.74 (m, 2H), 1.13–0.94 (m, 7H), 0.52 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 170.0, 149.2, 148.3, 139.9, 138.9, 137.2, 135.2, 130.4, 127.0, 126.7, 126.4, 125.6, 125.2, 125.1, 122.9, 122.5, 120.4, 120.4, 120.2, 120.0, 117.8, 114.3, 114.0, 112.8, 111.0, 110.9, 108.4, 98.5, 85.1, 84.6, 58.3, 55.9, 55.8, 54.0, 43.6, 41.7, 33.5, 29.2, 29.2, 28.7, 28.6, 22.9, 21.5; HRMS (FAB) m/z 728.3339 [calcd for $C_{44}H_{46}N_3O_7$ (M+H) 728.3336].
- 14. (-)-Indolocarbazole 16 (isomer 2): a pale yellow powder; $[\alpha]_{D}^{20}$ -30.0 (c 0.26, CHCl₃); IR (thin film/NaCl) 3485 (br m), 3334 (br w), 3056 (m), 2998 (m), 2928 (s), 2854 (m), 1732 (s), 1671 (s), 1585 (m), 1515 (s), 1487 (m), 1450 (s), 1400 (s) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.76 (d, J=8.0 Hz, 1H), 8.16 (d, J=8, 1 Hz, 1H), 7.94 (d, J=8.5 Hz, 1H), 7.88 (d, J=8.5 Hz, 1H), 7.51 (app t, J=7.8 Hz, 1H), 7.44 (app t, J=7.7 Hz, 1H), 7.37-7.26 (m, 2H), 7.15 (dd, J=5.0, 7.3 Hz, 1H), 7.11 (d, J=1.8Hz, 1H), 7.02 (dd, J=1.8, 8.1 Hz, 1H), 6.92 (d, J=8.1Hz, 1H), 5.57 (ddt, J = 10.2, 17.0, 6.8 Hz, 1H), 5.39 (s, 1H), 5.32 (d, J=15.1 Hz, 1H), 5.32 (app t, J=3.5 Hz, 1H), 4.78 (m, 1H), 4.75 (m, 1H), 4.43 (d, J=15.1 Hz, 1H), 4.00 (s, 3H), 3.76 (s, 3H), 3.75 (s, 3H), 3.54 (dd, J=7.3, 13.8 Hz, 1H), 2.56 (m, 1H), 2.47–2.33 (m, 2H), 2.18 (s, 3H), 1.79–1.67 (m, 2H), 1.07–0.87 (m, 7H), 0.45 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 170.0, 149.3, 148.3, 140.0, 138.9, 137.4, 135.1, 130.4, 127.1, 126.6, 126.6, 125.7, 125.3, 125.2, 122.7, 122.6, 120.6, 120.4, 120.2, 120.0, 118.1, 114.4, 114.0, 113.0, 111.1, 111.0, 108.3, 98.6, 85.1, 84.3, 58.3, 55.9, 55.9, 54.0, 43.6,

42.1, 33.5, 29.2, 28.9, 28.7, 28.6, 22.8, 21.5; HRMS (FAB) m/z 728.3340 [calcd for C₄₄H₄₆N₃O₇ (M+H) 728.3336].

- Schwab, P.; Grubbs, R. H.; Ziller, J. W. J. Am. Chem. Soc. 1996, 118, 100–110.
- 16. Due to signal overlap in ¹H NMR, E/Z ratio was not determined.
- 17. (-)-(7S)-K252a dimer (2): a white powder: $[\alpha]_{D}^{20}$ -145 (c 0.10, CHCl₃); IR (thin film/NaCl) 3223 (br m), 3051 (m), 2924 (s), 2851 (m), 1731 (s), 1678 (s), 1583 (m), 1487 (m), 1453 (s), 1392 (s) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.37 (d, J=7.8 Hz, 2H), 8.13 (d, J=7.8 Hz, 2H), 8.10 (s, 2H), 8.00 (d, J=8.5 Hz, 2H), 7.74 (d, J=8.3 Hz, 2H), 7.48–7.40 (m, 4H), 7.34 (app t, J=7.4 Hz, 2H), 7.26 (app t, J=7.6 Hz, 2H), 7.12 (dd, J=4.8, 7.4 Hz, 2H), 5.38 (s, 2H), 5.33 (dd, J=2.2, 7.8 Hz, 2H), 4.00 (s, 6H), 3.49 (dd, J = 7.4, 14.0 Hz, 2H), 2.51 (m, 2H), 2.30 (dd, J = 4.8, 14.0 Hz, 2H), 2.20 (s, 6H), 1.74 (m, 2H), 1.61 (m, 2H), 1.47–0.99 (m, 22H); 13 C NMR (100 MHz, CDCl₃): δ 173.3, 171.1, 140.7, 137.0, 136.7, 129.3, 125.4, 125.3, 124.8, 123.7, 121.1, 120.8, 120.4, 118.7, 116.2, 115.9, 115.8, 113.4, 107.0, 100.0, 86.2, 85.0, 57.2, 53.1, 42.6, 34.2, 29.3, 29.2, 29.1, 28.7, 28.7, 26.6, 22.7; HRMS (FAB) m/z 1129.5073 [calcd for $C_{68}H_{69}N_6O_{10}$ (M+H) 1129.5075].
- 18. (+)-(7*R*)-K252a Dimer (3): a white powder: $[\alpha]_{D}^{20}$ +107 (c 0.10, CHCl₃); IR (thin film/NaCl) 3499 (w), 3405 (br m), 3205 (br m), 3053 (m), 2924 (s), 2853 (s), 1732 (s), 1680 (s), 1584 (m), 1488 (w), 1458 (s), 1393 (s) cm⁻¹; ¹H NMR (400 MHz, acetone- d_6): δ 9.42 (d, J = 8.0 Hz, 2H), 8.17 (s, 2H), 8.15 (d, J=8.3 Hz, 2H), 8.01 (d, J=8.4 Hz, 2H), 7.78 (d, J = 8.3 Hz, 2H), 7.48 (app t, J = 7.8 Hz, 2H), 7.44 (app t, J=7.9 Hz, 2H), 7.34 (app t, J=7.3 Hz, 2H), 7.28 (app t, J=7.6 Hz, 2H), 7.15 (dd, J=5.1, 7.3 Hz, 2H), 5.46 (s, 2H), 5.38 (dd, J=1.9, 8.1 Hz, 2H), 4.00 (s, 6H), 3.52 (dd, J=7.3, 14.3 Hz, 2H), 2.56 (m, 2H), 2.34 (dd, J = 5.1, 14.3 Hz, 2H), 2.20 (s, 6H), 1.74–1.55 (m, 6H), 1.53–1.06 (m, 20H); ¹³C NMR (100 MHz, CDCl₃): δ 174.0, 172.4, 140.2, 137.8, 137.1, 129.1, 126.8, 125.7, 124.9, 124.4, 124.2, 123.3, 122.1, 120.6, 120.1, 117.1, 115.0, 114.1, 107.3, 98.7, 85.1, 84.4, 57.7, 53.9, 41.9, 34.3, 29.3, 29.3, 29.2, 29.2, 29.2, 26.4, 22.9; HRMS (FAB) m/z 1129.5073 [calcd for C₆₈H₆₉N₆O₁₀ (M+H) 1129.5075].